

1 **REVIEW**

2 **SPECIAL ISSUE ON INVASIVE MAMMAL SPECIES**

3

4 **Genetic tools in the management of invasive mammals: recent trends and**
5 **future perspectives**

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26 **ABSTRACT**

- 27 1. Invasive non-native species are now considered to be one of the greatest
28 threats to biodiversity worldwide. Therefore, efficient and cost-effective
29 management of species invasions requires robust knowledge of their
30 demography, ecology and impacts, and genetic-based techniques are
31 becoming more widely adopted in acquiring such knowledge.
- 32 2. We focus on the use of genetic tools in the applied management of
33 mammalian invasions globally, as well as on their inherent advantages and
34 disadvantages. We cover tools that are used in: (1) detecting and monitoring
35 mammalian invaders; (2) identifying origins and invasive pathways; (3)
36 assessing and quantifying the negative impacts of invaders; and 4) population
37 management and potential eradication of invasive mammals.
- 38 3. We highlight changes in sequencing technologies, including how the use of
39 techniques such as Sanger sequencing and microsatellite genotyping, for
40 monitoring and tracing invasive pathways respectively, are now giving way to
41 the use of high-throughput sequencing methods. These include the
42 emergence of environmental DNA (eDNA) metabarcoding for the early
43 detection of invasive mammals, and single nucleotide polymorphisms or
44 whole genomes to trace the sources of invasive populations. We are now
45 moving towards trials of genome-editing techniques and gene drives to control
46 or eradicate invasive rodents.
- 47 4. Genetic tools can provide vital information that may not be accessible with
48 non-genetic methods, for the implementation of conservation policies (e.g.
49 early detection using systematic eDNA surveillance, the identification of novel
50 pathogens). However, the lack of clear communication of novel genetic

51 methods and results (including transparency and reproducibility) to relevant
52 stakeholders can be prohibitive in translating these findings to appropriate
53 management actions. Geneticists should engage early with stakeholders to
54 co-design experiments in relation to management goals for invasive
55 mammals.

56 INTRODUCTION

57 The introduction of species outside of their native range has escalated due to
58 increased movement of people (Hulme 2009), and invasive non-native species are
59 now considered to be one of the greatest threats to biodiversity worldwide (Bellard et
60 al. 2016). Invasive species disrupt ecosystem services and lead to the introduction of
61 novel diseases, ultimately impacting native wildlife, domesticated species and
62 humans. In response to invasive non-native species, plans and policies are put into
63 place to prevent their entry and reduce or eliminate their impact. Such measures are
64 extremely costly in economic terms. For example, the European Union alone spends
65 approximately €12 billion annually on the control and management of invasive non-
66 native species and on mitigating their adverse impacts.

67

68 Efficient and cost-effective management of species invasions requires robust
69 knowledge of their demography, ecology and impacts, and genetic-based techniques
70 are becoming more widely adopted in acquiring such knowledge (Searle 2008,
71 Darling et al. 2017). The genetic tools that we have to study these processes have
72 developed dramatically over time, particularly over the last decade, and have
73 become more affordable, efficient and available for small to medium-scale
74 laboratories, providing new opportunities to study multiple aspects of invasions (Lee
75 2002). However, genetic tools are variable in methodology, design, price, complexity
76 and the resolution of results. The scope of this review is to provide an accessible
77 synopsis of the genetic techniques for the non-geneticist in order to enable
78 stakeholders, such as state and conservation managers, policy-makers, field
79 biologists and early-career researchers, to work collaboratively with geneticists to
80 address questions related to the prevention and management of mammalian

81 invasions. We provide a brief overview of effective genetic techniques that are
82 available for four management stages of a mammalian invasion: (1) detection and
83 monitoring of non-native invasive mammals; (2) identifying invaders' origins and
84 invasive pathways; (3) assessing and quantifying the negative impacts of invaders;
85 and (4) population management and the potential eradication of invasive mammals.

86

87 **DETECTION AND MONITORING**

88 The early and rapid detection of newly introduced mammals is vital to prevent further
89 spread that could subsequently result in a more costly eradication programme. Given
90 the elusive nature of many mammalian species, detection and monitoring often
91 requires indirect observations such as searching for latrines, faeces, hair, or tracks,
92 or direct observations such as live-trapping or camera-trapping surveys (Sales et al.
93 2020a). These can require differing levels of expertise and resources, but despite
94 high levels of expertise it is not always possible to assign indirect field signs correctly
95 to a species without further confirmation via DNA analysis (Harrington et al. 2010).

96

97 Indirect field signs such as hair and faeces can be subjected to genetic non-invasive
98 sampling (gNIS; Ferreira et al. 2018) to confirm species identification. gNIS has the
99 benefit of collecting genetic information without handling animals, which may cause
100 stress. Routine **PCR** (terms in **bold** are defined in the Glossary) methodologies can
101 be applied as diagnostic tools for identifying species from ambiguous field signs such
102 as hair or faeces. The required species-specific primers are already available to
103 identify, for example, Iberian carnivores from faecal DNA, including invasive
104 mammals such as the genet *Genetta genetta*, Egyptian Mongoose *Herpestes*
105 *ichneumon* and the North American mink *Neovison vison* (Fernandes et al. 2008).

106

107 DNA obtained from gNIS may have degraded into smaller fragments due to
108 prolonged exposure to environmental factors such as temperature fluctuations and
109 ultraviolet light. Therefore, PCR detection or identification methods can be used to
110 target short genetic regions (<1000 base pairs). **qPCR** is marginally more complex
111 but has some benefits over traditional PCR for the identification of species from
112 gNIS. qPCR can amplify shorter DNA regions (<100 base pairs) and is more
113 sensitive to smaller starting amounts of DNA. It has been used to detect invasive
114 mammals such as the greater white-toothed shrew *Crocidura russula* and grey
115 squirrel *Sciurus carolinensis* from native pine marten *Martes martes* faeces (O'Meara
116 et al. 2014). qPCR has the additional benefit of providing quality control to select
117 optimal DNA samples for further analysis, such as sequencing and genotyping, thus
118 allowing researchers to avoid wasting resources on poor-quality samples that are
119 unlikely to yield results. Kierepka et al. (2016) used qPCR to screen feral pig *Sus*
120 *scrofa* faecal-derived DNA prior to genotyping, to generate a robust capture-mark-
121 recapture protocol in order to facilitate accurate estimates of abundance.

122

123 Physical samples such as faeces or hair are not always required for species
124 detection. Organisms leave genetic material behind in the surrounding environment
125 (e.g. in water bodies and soil) via excretions and secretions (Harper et al. 2019); this
126 is referred to as **environmental DNA (eDNA)**. Single-species detection from eDNA
127 is possible using PCR, qPCR or droplet digital PCR (**ddPCR**). Research on invasive
128 wild boar *Sus scrofa* in North America (Williams et al. 2018) has demonstrated the
129 efficiency of a species-specific qPCR approach on samples from various water
130 bodies in detecting the species, but has also highlighted that a minimum number of

131 individuals is required for detection. This clearly has implications for providing early
132 detection of invasive species, which may initially be present in low numbers.

133

134 Single-species detection methods are relatively cheap, fast and robust, but require
135 prior knowledge of the target species to design appropriate detection methods (e.g.
136 O'Meara et al. 2014). If prior knowledge of the target species is unavailable, species
137 can be identified from gNIS using **Sanger sequencing** to generate a **DNA barcode**
138 (Hebert et al. 2003). In the Scottish Highlands, UK, experienced field surveyors used
139 field signs such as faeces to identify 57 sites out of 147 as positive for the presence
140 of invasive North American mink. Subsequent DNA sequencing of a standardised
141 portion of **mitochondrial DNA** (mtDNA) showed that mink faeces were misidentified
142 at all sites, and that they were commonly confused with native carnivore faeces
143 (Harrington et al. 2010). Had management or eradication programmes been
144 designed based on indirect observation, the result would have been a costly, time-
145 consuming, and unnecessary eradication programme.

146

147 **Next-generation sequencing** can facilitate the simultaneous identification of entire
148 communities (i.e. multiple species). **DNA metabarcoding** from environmental
149 samples has the potential to be used as an early warning system for the detection of
150 invasive non-native species, can be used for continuous monitoring programmes,
151 and has been extensively applied for tracking biological invasions in aquatic
152 ecosystems (Deiner et al. 2017). eDNA metabarcoding studies targeting mammalian
153 communities were relatively rare in comparison to other taxonomic groups (Sales et
154 al. 2020a), but this may change now that there are established metabarcoding

155 protocols for detecting and monitoring whole communities using vertebrate (Harper
156 et al. 2019) or mammal-specific primer sets (Ushio et al. 2017, Sales et al. 2020a,b).

157

158 eDNA metabarcoding is an emerging technique for invasive mammal detection and
159 monitoring, and there are important considerations for its use. For example,
160 mammals with larger home ranges (e.g. invasive carnivores) have lower probabilities
161 of detection than more abundant group-living mammals (Harper et al. 2019, Sales et
162 al. 2020a). Due to the high sensitivity of metabarcoding, contamination is a concern
163 (Sales et al. 2020a). It is therefore essential that specialised eDNA lab facilities (akin
164 to working with ancient DNA) are used (Zinger et al. 2019). Another consideration is
165 the existence of gaps in customised or online **reference databases** for identifying
166 sequences to the appropriate species level in under-studied geographic regions
167 (Sales et al. 2020b). However, with a carefully planned experimental design and the
168 appropriate field and lab controls (Zinger et al. 2019), eDNA metabarcoding has the
169 potential to be applied for early detection and ongoing surveillance of invasive
170 mammals (Harper et al. 2019, Sales et al. 2020a).

171

172 **ORIGINS AND INVASIVE PATHWAYS**

173 Identifying the origins of invasions is a critical management strategy in controlling the
174 spread of invasive species (Hulme 2009). When there is an absence of direct
175 evidence indicating the routes of invasion (such as records from interception at
176 ports), indirect methods such as the analyses of genetic data from invasive
177 populations and putative sources becomes vital (Searle 2008, Gargan et al. 2016).

178

179 Studies initially relied upon sequencing mtDNA to track the transport of invasive
180 species, because of the availability of universal primers for mammals (for mtDNA
181 genes such as cytochrome *b* and the control region) and available sequences (from
182 the native ranges for comparisons) in reference databases such as Genbank. For
183 invasive mammals with a global distribution, such as house mice *Mus* spp. and rats
184 *Rattus* spp., phylogenetic analyses of mtDNA have proven extremely useful in
185 tracing multiple introductions to islands and different continents over recent millennia
186 and centuries (Jones et al. 2013). The use of this type of mtDNA marker can be
187 limited over the spatial and temporal scales required for tracking more recent
188 invasions. Although mtDNA accumulates substitutions more rapidly than nuclear
189 DNA, mtDNA markers are generally useful for investigating intraspecific relationships
190 over tens to hundreds of thousands of years. Unless mtDNA variation is sufficiently
191 high in the native range, it is not ideal for tracing most mammalian invasions (Gray et
192 al. 2014) and may reveal the continent of origin as opposed to the country (Gargan
193 et al. 2016). Raccoons *Procyon lotor* show limited mtDNA diversity within their
194 invasive range in Europe, which originally led researchers to believe that they were
195 descended from a small number of founding individuals (Frantz et al. 2013).
196 However, the analysis of more rapidly evolving **microsatellites** led to the conclusion
197 that there were potentially up to four separate sources for the raccoon's current
198 distribution within its invasive range (Fischer et al. 2015). In the same vein, studies of
199 house mice have revealed the importance of using a multiple marker approach (such
200 as microsatellites) when inferring the origins of island populations, as many display
201 admixed origins (e.g. Gray et al. 2014).

202

203

204 Given that we are now firmly entrenched within the genomics era of molecular
205 ecology research, it is unsurprising that studies inferring the origins of invasive
206 mammals are now switching to **Single Nucleotide Polymorphism (SNP)** marker-
207 based approaches. Compared to microsatellite markers, SNPs usually span across a
208 greater proportion of the genome, can determine population demographics to a finer
209 scale, and do not require calibration between laboratories (Iacolina et al. 2016).
210 Incorporating SNPs in a study previously required a huge investment of time and
211 resources, usually applied only to economically important species (e.g. cattle, dogs,
212 rodents, pigs). The *de novo* discovery of SNPs in non-model organisms is now
213 achievable and affordable through **reduced representation sequencing** techniques
214 (such as Restriction Site Associated sequencing or RAD-seq; Baird et al. 2008).

215

216 Puckett et al. (2016) used ~32000 SNPs (derived from ddRAD genotyping) to
217 examine the population genomic structure of brown rats *Rattus norvegicus*
218 throughout their worldwide geographic range. Brown rats were generally grouped
219 into Asian and non-Asian groups, but fine-scale structuring was identified within
220 regions, reflecting more recent invasion pathways. For example, mtDNA data
221 revealed a European origin for contemporary New Zealand and western USA
222 populations, but SNP data revealed ancestry from admixed Asian and non-Asian
223 genomic clusters. In tracking the invasion of raccoon dogs *Nyctereutes procyonoides*
224 in Denmark, Nørgaard et al. (2017) utilised genotyping-by-sequencing to identify
225 over 4000 SNPs to trace their origins to Danish fur farms and reveal subsequent
226 admixture with neighbouring German populations. Unlike with microsatellites, newly
227 generated data on finer spatial scales can be compared with a global dataset of
228 SNPs if a reference genome is available. For example, this allowed Combs et al.

229 (2018) to determine that the most likely origins of the New York, USA, population of
230 brown rats were France and the British Isles.

231

232 **NEGATIVE IMPACTS**

233 **Diet and competition**

234 Invasive mammals may affect local flora and fauna through predation or ingestion
235 (e.g. feral cats take terrestrial vertebrates; Doherty et al. 2017), or via increased
236 competition (e.g. invasive American mink compete with native European mammalian
237 carnivores Sidorovich et al. 2010). Mammals are notoriously elusive, making their
238 diet difficult to document through direct observations, so that morphological
239 diagnostics of prey remains from stomach contents and faeces are a popular method
240 (Brzeziński et al. 2018). This methodology produces biased results due to variable
241 degradation rates between species and body parts (i.e. soft body parts degrade
242 faster than hard body parts), and residual body fragments that are found are difficult
243 to identify to species level (Deagle et al. 2009). Stable isotope analysis shows
244 promise, but has difficulties identifying prey species when isotopic signatures
245 naturally vary between geographic locations (Chibowski et al. 2019).

246

247 Genetic tools require DNA to be extracted from faeces or gut contents using
248 appropriate extraction kits capable of removing inhibitors associated with the
249 digestive tract. Species-specific primers and PCR are straight-forward and cost-
250 effective methods to measure predation rates of a single species of interest
251 (Waraniak et al. 2018). However, invasive mammals can have a variable diet
252 between native and introduced ranges (Ballari & Barrios-García 2014), making it
253 difficult to predict what they will consume in their introduced range. DNA

254 metabarcoding is a promising method: it allows the identification of multiple dietary
255 components of hundreds of individuals, and increases prey detection from 2% using
256 morphological diagnostics to 70% using metabarcoding (Pompanon et al. 2012,
257 Egeter et al. 2015a).

258

259 Not only can DNA metabarcoding accurately document an animal's impact on local
260 resources, but it can also reduce ambiguity. Previous assessments of the impact of
261 invasive rats *Rattus rattus* on endemic amphibians in New Zealand relied on
262 abundance estimates of native frog species in comparison to arrival patterns of the
263 invasive rat (Egeter et al. 2015b). Inconsistencies between observers caused doubt,
264 but DNA metabarcoding clarified the rat's consumption of New Zealand's native frog
265 species and its contribution to the population declines (Egeter et al. 2019). The
266 sensitivity achieved from next-generation sequencing methods allows multiple prey
267 items to be identified to the species level and generates a comprehensive account of
268 multiple animals' resource use and overlap. Telfair's skink *Leiopisma telfairii* was
269 introduced to Ile aux Aigrettes, Mauritius, Indian Ocean, for conservation purposes,
270 but unexpectedly met potential threats from the invasive Asian musk shrew *Suncus*
271 *murinus*. Species-specific primers showed the two species did not predate one
272 another (once adulthood was attained), but DNA metabarcoding identified significant
273 prey overlap and resulted in the suggestion that controlling shrew populations would
274 benefit the skink population (Brown et al. 2014).

275

276 Metabarcoding projects for dietary studies require some important considerations
277 before they are started (also relevant to eDNA metabarcoding studies, see above).
278 The first is targeting the appropriate genetic region for the target taxa in the diet,

279 such as vertebrates, invertebrates or plants (Kress et al. 2015). To know the full diet
280 of an omnivorous invader (e.g. wild boar), multiple regions are required for the full
281 taxonomic range within their diet (De Barba et al. 2014). Alternatively, highly
282 degenerative (non-specific) primers can be used to capture a wider range of prey
283 taxa, but this can result in over-representation of higher-quality host DNA (Zeale et
284 al. 2011). The broader the primers' taxonomic range, the more likely the chance of
285 amplifying non-target taxa and reducing the amount of information on a species' diet.
286 Blocking primers can mitigate host DNA amplification, but require more time to
287 design and test, as they may also block the amplification of some target prey taxa
288 (Su et al. 2018). The high sensitivity of PCR and high-throughput sequencing can
289 also result in the detection of taxa through secondary predation (i.e. detecting the
290 food of the food; Sheppard et al. 2005). Another difficulty is the inference of biomass
291 or the number of prey individuals from molecular diet analysis (Deagle et al. 2019).
292 Estimates of prey proportion are biased towards harder-bodied organisms due to
293 differential degradation rates. There are multiple ways to determine the importance
294 of certain taxa within a predator's diet, such as frequency of occurrence or relative
295 abundance (reviewed by Deagle et al. 2019).

296

297 **Disease**

298 The introduction of mammals into novel environments comes with the risk of co-
299 introducing pathogens or parasites that local fauna have not yet developed
300 resistance to (Paziewska et al. 2011). Mammalian invasions in Europe are likely to
301 have been responsible for the transport of pathogens responsible for salmonellosis,
302 toxoplasmosis and leptospirosis (Hulme 2014), and for the dissemination of the
303 plague across continents via rodent introductions (Gage & Kosoy 2005). Genetic

304 tools are becoming pivotal in disease management in wildlife (DeCandia et al. 2018):
305 PCR is currently used to verify morphological identification of pathogens and
306 parasites (Bagrade et al. 2016), and genetic tools can be used as detection methods
307 when there are difficulties in recreating optimal cell growing conditions to test for
308 prevalence levels (Heuser et al. 2017).

309

310 Different pathogen genotypes or strains can have different infection capabilities
311 (Nally et al. 2016). Sequencing actin genes of pathogenic *Cryptosporidium* revealed
312 that invasive raccoons harboured genotypes capable of infecting humans
313 (Leśniańska et al. 2016). To gain a higher resolution of bacterial population structure
314 and evolution, and to help understand the distribution of pathogenic species and
315 genotypes in novel areas invaded by mammalian hosts, multiple loci or genes can be
316 sequenced in multi-locus sequence typing (Margos et al. 2008). This method was
317 applied to *Borrelia* spp., an important pathogen in zoonotic ecology due to its
318 responsibility for Lyme disease. Sanger sequencing of the housekeeping gene (*clpA*)
319 and the infection-related gene (*ospC*) of *Borrelia burgdorferi* showed that invasive
320 grey squirrels in the UK are reservoirs for multiple *Borrelia burgdorferi* strains that
321 can affect multiple vertebrate clades (Millins et al. 2015). For larger-scale projects
322 and maximum efficiency, next-generation sequencing can be adapted for multi-locus
323 sequence typing from 100–200 samples in a cost-effective manner (Jacquot et al.
324 2014); this method was used to identify different *Borrelia* spp. lineages associated
325 with different small mammal host species (Jacquot et al. 2014).

326

327 Standardisation of sequence data is encouraged, and uploading data to online
328 databases allows combinations of multiple datasets to be incorporated into new and

329 broad meta-analyses (Maiden 2006). Phylogenetic analysis of openly available
330 sequence data from online reference databases allowed Hayman et al. (2013) to
331 decipher the origins, dissemination and diversification of the zoonotic pathogen
332 *Bartonella* spp. in mammalian clades and introductions.

333

334 **Hybridisation**

335 Hybridisation among species which are naturally separated is undoubtedly
336 increasing due to anthropogenic impacts, including species' invasions (McFarlane &
337 Pemberton 2019). Extensive introgression from invading populations can put already
338 endangered native populations at risk (Senn et al. 2019). Identifying hybrids based
339 on phenotypic characteristics is problematic due to intermediate phenotypes and
340 observer biases (McDevitt et al. 2009). To increase the efficiency of hybrid
341 identification, molecular markers have long been deployed; microsatellites have
342 been used since the 1990s. The increased use of assignment-based analysis in the
343 early 2000s (e.g. Randi et al. 2001) allowed researchers to identify the proportion of
344 the genome (usually inferring from ≥ 10 microsatellites) assigned to each species in
345 each individual, which individuals exhibited an admixed genotype and could
346 therefore be labelled as hybrids, and the percentage of the population consisting of
347 hybrids. This type of analyses has been important in providing initial indications of
348 the level of hybridisation between invasive sika deer *Cervus nippon* and red deer
349 *Cervus elaphus* in Europe (e.g. McDevitt et al. 2009), domestic/feral cats and
350 wildcats *Felis silvestris* in Europe (e.g. Randi et al. 2001) and domestic/feral pigs and
351 wild boar in multiple geographic regions (e.g. Scandura et al. 2011). While
352 microsatellite markers can be informative in detecting first or second-generation
353 hybridisation events, their low coverage means that they cannot detect extensive

354 backcrossing over several generations between parental species (McFarlane &
355 Pemberton 2019).

356

357 In order to improve resolution in detecting hybrids and their backcrosses, there is
358 clearly a need to use higher-density and diagnostic SNPs (Mattucci et al. 2019).
359 Several recent studies have highlighted improvements in hybrid detection by using
360 thousands of SNPs rather than 10 - 25 microsatellites. For example, a study on wolf-
361 dog hybridisation showed that only 1-5% of individuals were identified as hybrids
362 when 16 or 18 microsatellites were used (Randi 2008). A later study used 61000
363 SNPs to infer that 62% of Eurasian wolves *Canis lupus* had some level of admixture
364 with domestic dogs *Canis familiaris* (Pilot et al. 2018). In a well-studied hybrid zone
365 between sika deer and red deer in Kintyre, Scotland, an increased panel of 45000
366 SNPs reclassified 26% of individuals as hybrids that had originally been assigned to
367 one of the parental species from a previous study based on 22 microsatellites
368 (McFarlane et al. 2019). In attempting to preserve Scottish wildcats from extensive
369 introgression with feral/domestic cats, only wildcat individuals with both high genetic
370 scores (using a SNP panel) and high phenotype scores of wildcat 'purity' are
371 selected for the captive breeding and reintroduction programmes (Senn et al. 2019).

372

373 **MANAGEMENT AND ERADICATION**

374 Given the financial and social commitments required from stakeholders to undertake
375 long-term eradication programmes of species such as grey squirrels and American
376 mink, it is important to be able to gauge the success and impact of these efforts.
377 Microsatellites are very effective in determining recent changes in invasive mammal

378 population demographics in order to assess the progress of management and
379 control schemes (Velando et al. 2017).

380

381 Culling programs are well-established for the control of American mink in several
382 European countries. Fraser et al. (2013) used microsatellites to divide the Scottish
383 mink population into genetic clusters (sub-populations) which were classified as
384 management units. These units were formed through a combination of historical fur-
385 farm escapes and subsequent natural movement through a mosaic landscape
386 throughout Scotland. The genetic analysis of Scottish mink populations
387 corresponded to the habitat characteristics, and allowed Fraser et al. (2013) to
388 create an informed proposal on how to reduce the spread of the species and decide
389 where to direct eradication efforts. However, Oliver et al. (2016) used similar data to
390 identify a possible mechanism for populations in mainland Scotland remaining
391 relatively stable despite culling. They identified an increase in long-distance
392 immigration and an almost three-fold increase in male immigration into culled areas,
393 providing evidence of compensatory immigration during these culling efforts.

394

395 As with identifying invasive pathways with genetic markers, SNPs can provide higher
396 resolution of population demographics, and have been implemented instead of
397 traditional capture-mark-recapture methods to show connectivity and dispersal in
398 brown rat populations in an urban area (Combs et al. 2018). Piertney et al. (2016)
399 used 299 SNPs to identify genomic clusters of brown rats on the island of South
400 Georgia in order to identify the appropriate number of target areas for baiting
401 operations. Although these types of data (microsatellites and SNPs) and analyses
402 (population structure and gene flow) are useful for planning and assessing the

403 success of management and eradication programmes, an important consideration is
404 the likely response of the invader to control or eradication measures, whether these
405 be chemical or biological. For example, using a genome-wide SNPs, Morgan et al.
406 (2018) demonstrated that invasive house mice on islands off North and South
407 America did not possess rodenticide resistance alleles that are present in parts of
408 Europe (even though the study also found that these house mice were of European
409 ancestry). This has important implications for subsequent eradication and control
410 measures.

411

412 Even when genetic tools are used to identify key populations to target for culling
413 programs, culls require a lot of effort and usually only have the power to manage a
414 population rather than eradicate it completely. Island populations of invasive mice
415 have been proposed as targets for trialling more elaborate eradication programmes
416 involving genome-editing techniques using **CRISPR-Cas9** (Breed et al. 2019). The
417 concept of gene drives, whereby the use of genetic engineering alters the probability
418 of how specific alleles are inherited in future generations of offspring, is being tested
419 for eradication programmes in multiple invasive species, particularly invertebrates
420 (see Breed et al. 2019 for examples). Mammals are now being considered, and well-
421 studied model organisms such as mice are an obvious starting point. Transgenic
422 delivery of the male sex-determining factor (Sry) has been proposed to skew the sex
423 ratio heavily towards male mice and thereby control population size (Backus & Gross
424 2016). This would require repeated releases of engineered males, which could be
425 feasible on small islands (Campbell et al. 2015). Prowse et al. (2019) demonstrated
426 that the Y chromosome can be 'shredded' using CRISPR technology in mouse
427 embryonic stem cells, and individual-based simulations show that this targeted

428 deletion of a sex chromosome has the potential for eradicating an island population
429 of rodents. However, it would require >90% efficiency to produce high probabilities of
430 eradication success, and would be highly susceptible to changes in mating systems
431 and population size (Prowse et al. 2019).

432

433 There are additional concerns if genetically altered individuals are ever accidentally
434 released or spread beyond their target areas of control (a hallmark of effective
435 invaders), as gene drives are self-sustaining (Noble et al. 2018). Despite these
436 justifiable concerns, plans are already underway to bring this technique to the field
437 and to select an appropriate island for trials (Scudellari 2019). The use of the
438 technique is clearly complex in terms of scientific, social, regulatory and ethical
439 issues (Breed et al. 2019), and it remains to be determined how effective gene drives
440 will be over large geographic areas. However, gene drives offer a potentially more
441 targeted approach than the use of chemicals, which could impact non-target, native
442 species.

443

444 **CONCLUSION**

445 The impact and challenges of surveillance programmes related to invasive species
446 are recognised internationally and have led to the creation of policies aimed at
447 preventing and managing invasions. For example, the European Union has created
448 Regulation 1143/2014 on Invasive Alien Species (IAS Regulation) that contains three
449 measures to combat invasive species which include: (1) prevention, (2) early
450 detection and rapid eradication and (3) management. This review has highlighted
451 that genetic tools have multiple applications for the active management of invasive
452 mammalian species. Not only this, but they are reliable, robust, and provide vital

453 information, that may not be accessible with non-genetic methods, for the
454 implementation of conservation policies (e.g. early detection using systematic eDNA
455 surveillance and the identification of novel pathogens).

456

457 However, there are technical challenges associated with the standardisation of
458 genetic methodologies and bioinformatic pipelines used between laboratories, even
459 when researchers are attempting to address similar questions (Zinger et al. 2019).
460 For example, how the samples have been collected (e.g. gNIS or tissue) and stored
461 (e.g. in ethanol or frozen) has implications for what techniques can be performed
462 downstream in the laboratory (e.g. single gene or whole-genome approaches).
463 Another significant challenge is the availability of appropriate funding and expertise.
464 These factors can all limit what questions can be addressed that will translate into
465 management actions and decisions.

466

467 In addition, the lack of clear communication of novel genetic methods and results
468 (including transparency and reproducibility) to relevant stakeholders can be
469 prohibitive in translating these findings to appropriate management and eradication
470 action on the ground (Mosher et al. 2019, Ward et al. 2020). These communication
471 challenges have been well documented in relation to the marine sector (e.g. Darling
472 et al. 2017), but little coordination has taken place in relation to invasive mammals,
473 despite the environmental and economic consequences that invasive non-native
474 species pose to native species, habitats and the agricultural industry. Geneticists
475 should engage early with stakeholders in relation to project costs, duration and
476 management goals for invasive mammals. This will allow for robust experimental

477 design using existing genetic tools, and the development of new technologies that
478 can be tailored towards specific management issues (Mosher et al. 2019).

479

480 **REFERENCES**

- 481 Andrews KR, Good JM, Miller MR, Luikart G, Hohenlohe PA (2016) Harnessing the
482 power of RADseq for ecological and evolutionary genomics. *Nature Reviews*
483 *Genetics* 17: 81-92.
- 484 Backus GA, Gross K (2016) Genetic engineering to eradicate invasive mice on
485 islands: modeling the efficiency and ecological impacts. *Ecosphere* 7: e01589.
- 486 Bagrade G, Dekšne G, Ozoliņa Z, Howlett SJ, Interisano M, Casulli A, Pozio E
487 (2016) *Echinococcus multilocularis* in foxes and raccoon dogs: an increasing
488 concern for Baltic countries. *Parasites and Vectors* 9: 615.
- 489 Baird NA, Etter PD, Atwood TS, Currey MC, Shiver AL, Lewis ZA et al. (2008) Rapid
490 SNP discovery and genetic mapping using sequenced RAD markers. *PLoS*
491 *ONE* 3: e3376.
- 492 Ballari SA, Barrios-García MN (2014) A review of wild boar *Sus scrofa* diet and
493 factors affecting food selection in native and introduced ranges. *Mammal*
494 *Review* 44: 124–134.
- 495 Bellard C, Cassey P, Blackburn TM (2016) Alien species as a driver of recent
496 extinctions. *Biology Letters* 12: 20150623.
- 497 Breed MF, Harrison PA, Blyth C, Byrne M, Gaget V, Gellie NJC et al. (2019) The
498 potential of genomics for restoring ecosystems and biodiversity. *Nature Reviews*
499 *Genetics* 20: 615–628.
- 500 Brown DS, Burger R, Cole N, Vencatasamy D, Clare EL, Montazam A, Symondson
501 WOC (2014). Dietary competition between the alien Asian musk shrew (*Suncus*
502 *murinus*) and a re-introduced population of Telfair's skink (*Leiopisma telfairii*).
503 *Molecular Ecology* 23: 3695–3705.
- 504 Brzeziński M, Chibowska P, Zalewski A, Borowik T, Komar E (2018) Water vole

505 (*Arvicola amphibius*) population under the impact of the American mink
506 (*Neovison vison*): are small midfield ponds safe refuges against this invasive
507 predator? *Mammalian Biology* 93: 182–188.

508 Campbell KJ, Beek J, Eason CT, Glen AS, Godwin J, Gould F et al. (2015) The next
509 generation of rodent eradications: innovative technologies and tools to improve
510 species specificity and increase their feasibility on islands. *Biological
511 Conservation* 185: 47–58.

512 Chibowski P, Zalewski A, Suska-Malawska M, Brzeziński M (2019) Study on
513 geographical differences in American mink diets reveals variations in isotopic
514 composition of potential mink prey. *Mammal Research* 64: 343-351.

515 Combs M, Puckett EE, Richardson J, Mims D, Munshi-South J (2018) Spatial
516 population genomics of the brown rat (*Rattus norvegicus*) in New York City.
517 *Molecular Ecology* 27: 83–98.

518 Darling JA, Galil BS, Carvalho GR, Rius M, Viard F, Piraino S (2017)
519 Recommendations for developing and applying genetic tools to assess and
520 manage biological invasions in marine ecosystems. *Marine Policy* 85: 54–64.

521 De Barba M, Miquel C, Boyer F, Mercier C, Rioux D, Coissac E, Taberlet P (2014)
522 DNA metabarcoding multiplexing and validation of data accuracy for diet
523 assessment: application to omnivorous diet. *Molecular Ecology Resources* 14:
524 306–323.

525 Deagle BE, Clarke LJ, Thomas AC, McInnes JC, Vesterinen EJ, Clare EL et al.
526 (2019) Counting with DNA in metabarcoding studies: how should we convert
527 sequence reads to dietary data? *Molecular Ecology* 28: 391-406.

528 Deagle BE, Kirkwood R, Jarman SN (2009) Analysis of Australian fur seal diet by
529 pyrosequencing prey DNA in faeces. *Molecular Ecology* 18: 2022–2038.

530 DeCandia AL, Dobson AP, vonHoldt BM (2018) Toward an integrative molecular
531 approach to wildlife disease. *Conservation Biology* 32: 798–807.

532 Deiner K, Bik HM, Elvira M, Seymour M, Lacoursière-Roussel A, Altermatt F et al.
533 (2017) Environmental DNA metabarcoding: transforming how we survey animal
534 and plant communities. *Molecular Ecology* 26: 5872–5895.

535 Doherty TS, Dickman CR, Johnson CN, Legge SM, Ritchie EG, Woinarski JCZ
536 (2017) Impacts and management of feral cats (*Felis catus*) in Australia. *Mammal
537 Review* 47: 83–97.

538 Egeter B, Bishop PJ, Robertson BC (2015a) Detecting frogs as prey in the diets of
539 introduced mammals: a comparison between morphological and DNA-based
540 diet analyses. *Molecular Ecology Resources* 15: 306–316.

541 Egeter B, Robertson BC, Bishop PJ (2015b) A synthesis of direct evidence of
542 predation on amphibians in New Zealand, in the context of global invasion
543 biology. *Herpetological Review* 46: 512–519.

544 Egeter B, Roe C, Peixoto S, Puppo P, Easton LJ, Pinto J et al. (2019) Using
545 molecular diet analysis to inform invasive species management: a case study of
546 introduced rats consuming endemic New Zealand frogs. *Ecology and Evolution*
547 9: 5032-5048.

548 Fernandes CA, Ginja C, Pereira I, Tenreiro R, Bruford MW, Santos-Reis M (2008)
549 Species-specific mitochondrial DNA markers for identification of non-invasive
550 samples from sympatric carnivores in the Iberian Peninsula. *Conservation
551 Genetics* 9: 681–690.

552 Ferreira CM, Sabino-Marques H, Paupério J, Barbosa S, Costa P, Encarnação C et
553 al. (2018) Genetic non-invasive sampling (gNIS) as a cost-effective tool for
554 monitoring elusive small mammals. *European Journal of Wildlife Research* 64:

555 46.

556 Fischer ML, Hochkirch A, Heddergott M, Schulze C, Anheyer-Behmenburg HE, Lang
557 J et al. (2015) Historical invasion records can be misleading: genetic evidence
558 for multiple introductions of invasive raccoons (*Procyon lotor*) in Germany. *PLoS*
559 *ONE* 10: e0125441.

560 Frantz AC, Heddergott M, Lang J, Schulze C, Ansorge H, Runge M et al. (2013)
561 Limited mitochondrial DNA diversity is indicative of a small number of founders
562 of the German raccoon (*Procyon lotor*) population. *European Journal of Wildlife*
563 *Research* 59: 665–674.

564 Fraser EJ, Macdonald DW, Oliver MK, Piertney S, Lambin X (2013) Using population
565 genetic structure of an invasive mammal to target control efforts – an example of
566 the American mink in Scotland. *Biological Conservation* 167: 35–42.

567 Gage KL, Kosoy MY (2005) Natural history of the plague: perspectives from more
568 than a century of research. *Annual Review of Entomology* 50: 505–528.

569 Gargan LM, Cornette R, Yearsley JM, Montgomery WI, Paupério J, Alves PC et al.
570 (2016) Molecular and morphological insights into the origin of the invasive
571 greater white-toothed shrew (*Crocidura russula*) in Ireland. *Biological Invasions*
572 18: 857–871.

573 Gray MM, Wegmann D, Haasl RJ, White MA, Gabriel SI, Searle JB et al. (2014)
574 Demographic history of a recent invasion of house mice on the isolated island of
575 Gough. *Molecular Ecology* 23: 1923–1939.

576 Harper LR, Handley LL, Carpenter AI, Ghazali M, Di Muri C, Macgregor CJ et al.
577 (2019) Environmental DNA (eDNA) metabarcoding of pond water as a tool to
578 survey conservation and management priority mammals. *Biological*
579 *Conservation* 238: 108225.

580 Harrington LA, Harrington AL, Hughes J, Stirling D, Macdonald DW (2010) The
581 accuracy of scat identification in distribution surveys: American mink, (*Neovison*
582 *vison*), in the northern highlands of Scotland. *European Journal of Wildlife*
583 *Research* 56: 377–384.

584 Hayman DTS, McDonald KD, Kosoy MY (2013) Evolutionary history of rat-borne
585 *Bartonella*: the importance of commensal rats in the dissemination of bacterial
586 infections globally. *Ecology and Evolution* 3: 3195–3203.

587 Hebert PDN, Cywinska A, Ball SL, DeWaard JR (2003) Biological identifications
588 through DNA barcodes. *Proceedings of the Royal Society of London. Series B:*
589 *Biological Sciences* 270: 313–321.

590 Heuser E, Fischer S, Ryll R, Mayer-Scholl A, Hoffmann D, Spahr C et al. (2017)
591 Survey for zoonotic pathogens in Norway rat populations from Europe. *Pest*
592 *Management Science* 73: 341–348.

593 Hulme PE (2009) Trade, transport and trouble: managing invasive species pathways
594 in an era of globalization. *Journal of Applied Ecology* 46: 10–18.

595 Hulme PE (2014) Invasive species challenge the global response to emerging
596 diseases. *Trends in Parasitology* 30: 267–270.

597 Iacolina L, Scandura M, Goedbloed DJ, Alexandri P, Crooijmans RPMA, Larson G et
598 al. (2016) Genomic diversity and differentiation of a managed island wild boar
599 population. *Heredity* 116: 60–67.

600 Jacquot M, Bisseux M, Abrial D, Marsot M, Ferquel E, Chapuis JL et al. (2014) High-
601 throughput sequence typing reveals genetic differentiation and host
602 specialization among populations of the *Borrelia burgdorferi* species complex
603 that infect rodents. *PLoS ONE* 9: e88581

604 Jones EP, Eager HM, Gabriel SI, Jóhannesdóttir F, Searle JB (2013) Genetic

605 tracking of mice and other bioproxies to infer human history. *Trends in Genetics*
606 29: 298–308.

607 Kierepka EM, Unger SD, Keiter DA, Beasley JC, Rhodes Jr OE, Cunningham FL,
608 Piaggio AJ (2016) Identification of robust microsatellite markers for wild pig fecal
609 DNA. *Journal of Wildlife Management* 80: 1120–1128.

610 Kress WJ, García-Robledo C, Uriarte M, Erickson DL (2015) DNA barcodes for
611 ecology, evolution, and conservation. *Trends in Ecology and Evolution* 30: 25–
612 35.

613 Lee CE (2002) Evolutionary genetics of invasive species. *Trends in Ecology and*
614 *Evolution* 17: 386–391.

615 Leśniańska K, Perec-Matysiak A, Hildebrand J, Buńkowska-Gawlik K, Piróg A,
616 Popiołek M (2016). *Cryptosporidium* spp. and *Enterocytozoon bieneusi* in
617 introduced raccoons (*Procyon lotor*)—first evidence from Poland and Germany.
618 *Parasitology Research* 115: 4535–4541.

619 Maiden MCJ (2006) Multilocus sequence typing of bacteria. *Annual Review of*
620 *Microbiology* 60: 561–588.

621 Margos G, Gatewood AG, Aanensen DM, Hanincova K, Terekhova D, Vollmer SA et
622 al. (2008) MLST of housekeeping genes captures geographic population
623 structure and suggests a European origin of *Borrelia burgdorferi*. *Proceedings of*
624 *the National Academy of Sciences* 105: 8730–8735.

625 Mattucci F, Galaverni M, Lyons LA, Alves PC, Randi E (2019) Genomic approaches
626 to identify hybrids and estimate admixture times in European wildcat
627 populations. *Scientific Reports* 9: 11612.

628 McDevitt AD, Edwards CJ, O'Toole P, O'Sullivan P, O'Reilly C, Carden RF (2009)
629 Genetic structure of, and hybridisation between, red (*Cervus elaphus*) and sika

630 (*Cervus nippon*) deer in Ireland. *Mammalian Biology* 74: 263–273.

631 McFarlane SE, Hunter DC, Senn HV, Smith SL, Holland R, Huisman J, Pemberton
632 JM (2019) Increased genetic marker density reveals high levels of admixture
633 between red deer and introduced Japanese sika in Kintyre, Scotland.
634 *Evolutionary Applications* doi: 10.1111/eva.12880.

635 McFarlane SE, Pemberton JM (2019) Detecting the true extent of introgression
636 during anthropogenic hybridization. *Trends in Ecology and Evolution* 34: 315-
637 326.

638 Millins C, Magierecka A, Gilbert L, Edoff A, Brereton A, Kilbride E et al. (2015) An
639 invasive mammal (the gray squirrel, *Sciurus carolinensis*) commonly hosts
640 diverse and atypical genotypes of the zoonotic pathogen *Borrelia burgdorferi*
641 sensu lato. *Applied and Environmental Microbiology* 81: 4236–4245.

642 Morgan AP, Didion JP, Hughes JJ, Searle JB, Jolley WJ, Campbell KJ et al. (2018)
643 Genetic characterization of invasive house mouse populations on small islands.
644 *BioRxiv*, 332064.

645 Mosher BA, Bernard RF, Lorch JM, Miller DAW, Richgels KLD, White CL, Grant
646 EHC (2019) Successful molecular detection studies require clear
647 communication among diverse research partners. *Frontiers in Ecology and the*
648 *Environment* doi: 10.1002/fee.2141

649 Nally JE, Arent Z, Bayles DO, Hornsby RL, Gilmore C, Regan S et al. (2016)
650 Emerging infectious disease implications of invasive mammalian species: the
651 greater white-toothed shrew (*Crocidura russula*) is associated with a novel
652 serovar of pathogenic leptospira in Ireland. *PLoS Neglected Tropical Diseases*
653 10: e0005174.

654 Noble C, Adlam B, Church GM, Esvelt KM, Nowak MA (2018) Current CRISPR gene

655 drive systems are likely to be highly invasive in wild populations. *eLife* 7:
656 e33423.

657 Nørgaard LS, Mikkelsen DMG, Elmeros M, Chriél M, Madsen AB, Nielsen JL et al.
658 (2017) Population genomics of the raccoon dog (*Nyctereutes procyonoides*) in
659 Denmark: insights into invasion history and population development. *Biological*
660 *Invasions* 19: 1637–1652.

661 O'Meara DB, Sheehy E, Turner PD, O'Mahony D, Harrington AP, Denman H et al.
662 (2014) Non-invasive multi-species monitoring: real-time PCR detection of small
663 mammal and squirrel prey DNA in pine marten (*Martes martes*) scats. *Acta*
664 *Theriologica* 59: 111–117.

665 Oliver MK, Piertney SB, Zalewski A, Melero Y, Lambin X (2016) The compensatory
666 potential of increased immigration following intensive American mink population
667 control is diluted by male-biased dispersal. *Biological Invasions* 18: 3047–3061.

668 Paziewska A, Harris PD, Zwolińska L, Bajer A, Siński E (2011) Recombination within
669 and between species of the alpha proteobacterium *Bartonella* infecting rodents.
670 *Microbial Ecology* 61: 134–145.

671 Piertney SB, Black A, Watt L, Christie D, Poncet S, Collins MA (2016) Resolving
672 patterns of population genetic and phylogeographic structure to inform control
673 and eradication initiatives for brown rats *Rattus norvegicus* on South Georgia.
674 *Journal of Applied Ecology* 53: 332–339.

675 Pilot M, Greco C, VonHoldt BM, Randi E, Jędrzejewski W, Sidorovich VE et al.
676 (2018) Widespread, long-term admixture between grey wolves and domestic
677 dogs across Eurasia and its implications for the conservation status of hybrids.
678 *Evolutionary Applications* 11: 662–680.

679 Pompanon F, Deagle BE, Symondson WOC, Brown DS, Jarman SN, Taberlet P

680 (2012) Who is eating what: diet assessment using next generation sequencing.
681 *Molecular Ecology* 21: 1931–1950.

682 Prowse TAA, Adikusuma F, Cassey P, Thomas P, Ross JV (2019) A Y-chromosome
683 shredding gene drive for controlling pest vertebrate populations. *eLife* 8:
684 e41873.

685 Puckett EE, Park J, Combs M, Blum MJ, Bryant JE, Caccone A et al. (2016) Global
686 population divergence and admixture of the brown rat (*Rattus norvegicus*).
687 *Proceedings of the Royal Society B: Biological Sciences* 283: 20161762.

688 Randi E (2008) Detecting hybridization between wild species and their domesticated
689 relatives. *Molecular Ecology* 17: 285–293.

690 Randi E, Pierpaoli M, Beaumont M, Ragni B, Sforzi A (2001) Genetic identification of
691 wild and domestic cats (*Felis silvestris*) and their hybrids using Bayesian
692 clustering methods. *Molecular Biology and Evolution* 18: 1679–1693.

693 Sales NG, Mckenzie MB, Drake J, Harper LR, Browett SS, Coscia I et al. (2020a)
694 Fishing for mammals: landscape-level monitoring of terrestrial and semi-aquatic
695 communities using eDNA from lotic ecosystems. *Journal of Applied Ecology* in
696 press.

697 Sales NG, da Cruz Kaizer M, Coscia I, Perkins J, Highlands A, Boubli JP et al.
698 (2020b) Assessing the potential of environmental DNA metabarcoding for
699 monitoring Neotropical mammals: a case study in the Amazon and Atlantic
700 Forest, Brazil. *Mammal Review* doi: 10.1111/mam.12183.

701 Scandura M, Iacolina L, Apollonio M (2011) Genetic diversity in the European wild
702 boar *Sus scrofa*: phylogeography, population structure and wild x domestic
703 hybridization. *Mammal Review* 41: 125–137.

704 Scudellari M (2019) Self-destructing mosquitoes and sterilized rodents: the promise

705 of gene drives. *Nature* 571: 160–162.

706 Searle JB (2008) The genetics of mammalian invasions: a review. *Wildlife Research*
707 35: 185–192.

708 Senn HV, Ghazali M, Kaden J, Barclay D, Harrower B, Campbell RD et al. (2019)
709 Distinguishing the victim from the threat: SNP-based methods reveal the extent
710 of introgressive hybridization between wildcats and domestic cats in Scotland
711 and inform future in situ and ex situ management options for species restoration.
712 *Evolutionary Applications* 12: 399–414.

713 Sheppard SK, Bell J, Sunderland KD, Fenlon J, Skervin D, Symondson WOC (2005)
714 Detection of secondary predation by PCR analyses of the gut contents of
715 invertebrate generalist predators. *Molecular Ecology* 14: 4461–4468.

716 Sidorovich VE, Polozov AG, Zalewski A (2010) Food niche variation of European
717 and American mink during the American mink invasion in north-eastern Belarus.
718 *Biological Invasions* 12: 2207–2217.

719 Su M, Liu H, Liang X, Gui L, Zhang J (2018) Dietary analysis of marine fish species:
720 enhancing the detection of prey-specific DNA sequences via high-throughput
721 sequencing using blocking primers. *Estuaries and Coasts* 41: 560–571.

722 Ushio M, Fukuda H, Inoue T, Makoto K, Kishida O, Sato K et al. (2017)
723 Environmental DNA enables detection of terrestrial mammals from forest pond
724 water. *Molecular Ecology Resources* 17: e63–e75.

725 Velando A, Morán P, Romero R, Fernández J, Piorno V (2017) Invasion and
726 eradication of the American mink in the Atlantic Islands National Park (NW
727 Spain): a retrospective analysis. *Biological Invasions* 19: 1227–1241.

728 Waraniak JM, Blumstein DM, Scribner KT (2018) Barcoding PCR primers detect
729 larval lake sturgeon (*Acipenser fulvescens*) in diets of piscine predators.

730 *Conservation Genetics Resources* 10: 259–268.

731 Ward AI, Richardson S, Macarthur R, Mill AC (2020) Using and communicating
732 uncertainty for the effective control of invasive non-native species. *Mammal*
733 *Review* 50: in press.

734 Williams KE, Huyvaert KP, Vercauteren KC, Davis AJ, Piaggio AJ (2018) Detection
735 and persistence of environmental DNA from an invasive, terrestrial mammal.
736 *Ecology and Evolution* 8: 688–695.

737 Zeale MRK, Butlin RK, Barker GLA, Lees DC, Jones G (2011) Taxon-specific PCR
738 for DNA barcoding arthropod prey in bat faeces. *Molecular Ecology Resources*
739 11: 236–244.

740 Zinger L, Bonin A, Alsos IG, Bálint M, Bik H, Boyer F et al. (2019) DNA
741 metabarcoding—need for robust experimental designs to draw sound ecological
742 conclusions. *Molecular Ecology* 28: 1857–1862.

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745

746 **Glossary**

747 **CRISPR-Cas9**

748 A targeted genome-editing tool comprised of the programmable Cas9 endonuclease,
749 which introduces double-strand breaks into DNA; and a guide RNA, which targets
750 the Cas9 nuclease to a specific DNA sequence. This allows for a portion of a target
751 organism's genome to be modified by adding, removing or altering a DNA sequence.

752

753 **DNA barcode**

754 A DNA barcode is a standardised fragment of the genome that can be used to
755 identify a species. Cytochrome c oxidase I was traditionally the mtDNA marker of
756 choice in barcoding studies. The region is highly conserved throughout the animal
757 kingdom but is variable enough to differentiate between species (Hebert et al. 2003).

758

759 **DNA metabarcoding**

760 The use of universal primers to amplify multiple DNA barcodes from bulk samples
761 containing multiple species, such as stomach contents or environmental samples.

762

763 **Environmental DNA (eDNA)**

764 Extra-organismal DNA molecules that are shed in the environment. In animals,
765 eDNA can originate from skin, mucous, saliva, sperm, secretions, eggs, faeces,
766 urine and blood. eDNA can be used to detect the presence of species from samples
767 of soil, water, or other substances from the environment.

768

769 **Microsatellite**

770 Microsatellites are regions of nuclear DNA which have tandemly repeated regions.
771 These tandem repeats are generally 2–6 base pairs in length and have a very high
772 mutation rate. The variation of microsatellites between individuals and populations
773 can be used to determine population demographics such as gene flow, relatedness
774 and genetic diversity.

775

776 **Mitochondrial DNA**

777 Mitochondrial DNA (mtDNA), found in the mitochondria as opposed to in the nucleus,
778 has a number of favourable properties for phylogeographic and phylogenetic studies,
779 such as the absence of recombination (which results in an effectively clonal
780 inheritance from the maternal side) and a lack of both pseudogenes and repetitive
781 DNA. mtDNA tends to accumulate base pair substitutions at a higher rate than
782 nuclear DNA.

783

784 **Next-generation sequencing**

785 Next-generation sequencing, also known as high-throughput sequencing, is a broad
786 term used to describe a number of different modern sequencing technologies. A
787 large number of sequences (millions to billions of sequence reads) are generated on
788 a single sequencing run.

789

790 **PCR**

791 The **Polymerase Chain Reaction (PCR)** is the exponential amplification (i.e. makes
792 thousands of copies) of a specifically targeted region of DNA through repeated

793 heating and cooling cycles. It is an essential component in most genetic
794 methodologies as more copies of the region provides a stronger signal for
795 downstream analysis such as sequencing. Primers are required to target the region
796 of interest and can be designed to be species-specific or to work on a broad range of
797 species.

798

799 **qPCR**

800 qPCR is a process by which the DNA fragment is amplified like in normal PCR, but
801 the amplification rate of the DNA fragment is continuously monitored using
802 fluorescent light. The starting amount of DNA can then be quantified against a set of
803 known standards. Droplet digital PCR (**ddPCR**) does not monitor the amplification
804 process, but it can accurately quantify the starting amount of DNA without the
805 necessity for standards.

806

807 **Reduced representation sequencing**

808 In reduced representation sequencing, restriction enzymes are used to cut (digest)
809 the genome at specific cut sites, defined by a specific sequence of nucleotides.
810 Sequencing and clustering of these DNA fragments allows the *de novo* discovery of
811 SNPs. Variations of this method of sequencing include Restriction-site Associated
812 DNA sequencing (RAD-seq), double digest RAD sequencing (ddRAD) and
813 genotyping-by-sequencing (GBS). See Andrews et al. (2016) for a detailed review.

814

815 **Reference databases**

816 Generated DNA sequences and barcodes need to be compared to existing
817 sequences that have been identified as belonging to a species (or at least as a
818 genus, depending on the taxonomic group) by an expert. Reference databases
819 provide public access to such sequences. Examples include Genbank, the Barcode
820 of Life Database and the CDC Bartonella Laboratory database. Sequences in
821 reference databases should have been subjected to quality control for taxonomic
822 accuracy, but this is not always the case (particularly for older records).

823

824 **Sanger sequencing**

825 A region of DNA is copied using a fluorescent dye unique to each nucleotide. The
826 colours read by the machine can determine the sequence of nucleotides in the
827 region. Sanger sequencing is a low throughput method suited to sequencing long
828 strands (~1000 base pairs) of a single region of DNA.

829

830 **Single Nucleotide Polymorphism (SNP)**

831 These are single base pair changes/variations (polymorphisms) spanning across
832 hundreds to thousands of locations (loci) along the genome. Deciphering patterns of
833 these changes between multiple individuals can be used to determine population
834 demographics such as gene flow and levels of inbreeding. They can provide higher
835 resolution information compared to other genetic markers such as microsatellites. In
836 addition, they can be used to identify signatures of selection/adaptation in
837 populations.

838

839

840